MAINTENANCE OF HORMONE SENSITIVE PHOSPHOINOSITIDE POOLS IN THE PLASMA MEMBRANE REQUIRES PHOSPHATIDYLINOSITOL 4-KINASE III ALPHA.

Andras Balla[1,2], Yeun Ju Kim[1], Peter Varnai[1,2], Zsofia Szentpetery[1], Zachary Knight[3], Kevan M. Shokat[4] and Tamas Balla[1*]

[1] Endocrinology and Reproduction Research Branch, NICHD, National Institutes of Health, Bethesda, MD, USA, [2]Department of Physiology, Semmelweis University, School of Medicine, Budapest, Hungary. [3] Program in Chemistry and Chemical Biology, University of California, San Francisco, CA, USA, [4] Howard Hughes Medical Institute and Department of Cellular and Molecular Pharmacology, University of California, San Francisco, CA, USA

running title: regulation of hormone-sensitive phosphoinositide pools

Keywords: phosphoinositide, calcium, phosphatidylinositol 4-kinase, PH domain, green fluorescent protein, angiotensin II

*To whom correspondence should be addressed: National Institutes of Health, Bldg 49, Rm 6A35, 49 Convent Drive, Bethesda, MD 20892-4510, ph:(301)-496-2136, FAX, (301)-480-8010, E-mail: tambal@box-t.nih.gov

ABSTRACT

Type III phosphatidylinositol (PtdIns) 4-kinases (PI4Ks) have been previously shown to support plasma membrane phosphoinositide synthesis during phospholipase C activation and Ca^{2+} signaling. Here we use biochemical and imaging tools to monitor phosphoinositide changes in the plasma membrane in combination with pharmacological and genetic approaches to determine which of the type III PI4Ks (α or β) is responsible for supplying phosphoinositides during agonist-induced Ca^{2+} signaling. Using inhibitors that discriminate between the α- and β-isoforms of type III PI4Ks, PI4KIIIα was found indispensable for the production of PtdIns4*P*, PtdIns(4,5)P_2 and Ca^{2+} signaling in angiotensin II (AngII) stimulated cells. Down-regulation of either the type II or type III PI4K enzymes by siRNA, had small but significant effects on basal PtdIns4*P* and PtdIns(4,5)P_2 levels in ^{32}P-labeled cells but only PI4KIIIα downregulation caused a slight impairment of PtdIns4*P* and PtdIns(4,5)P_2 resynthesis in AngII stimulated cells. None of the PI4K siRNA treatments had a measurable effect on AngII induced Ca^{2+} signaling. These results indicate that a small fraction of the cellular PI4K activity is sufficient to maintain plasma membrane phosphoinositide pools and demonstrate the value of the pharmacological approach in revealing the pivotal role of PI4KIIIα enzyme in maintaining plasma membrane phosphoinositides.

INTRODUCTION

Activation of cell surface receptors by a variety of stimuli initiates a cascade of molecular events ultimately eliciting a response characteristic of the target cell. One of the most studied and best-characterized signal transduction pathways is initiated by the phospholipase C (PLC)–mediated breakdown of PtdIns(4,5)P_2 to generate the Ca^{2+}-mobilizing messenger, $InsP_3$ and the PKC activator, DAG (Berridge and Irvine, 1984). It has long been recognized that the sustained production of these messengers requires continuous phosphorylation of PtdIns to PtdIns4*P* and PtdIns(4,5)P_2 by PI 4-kinase and PIP 5-kinase enzymes, due to the limited amount of PtdIns(4,5)P_2 present in the plasma membrane (Creba *et al.*, 1983). Several isoforms within the PI 4-kinase and PIP 5-kinase families have been described (Doughman *et al.*, 2003; Balla and Balla, 2006), and a recent study identified a splice variant of PIP 5-kinase γ as the enzyme responsible for PtdIns4*P* to PtdIns(4,5)P_2 conversion in agonist-induced Ca^{2+} signaling (Wang *et al.*, 2004). In contrast, the PI 4-kinase that generates PtdIns4*P* for this process so far has eluded identification.

Four PI 4-kinase enzymes have been identified in mammalian cells that belong to either the type II or type III families (Balla and Balla, 2006). Type II PI 4-kinase enzymes (α- and β-forms) are small, 56 kDa proteins that are abundant in almost all cellular membranes and are enriched in plasma membrane preparations. They are kept in the membrane by palmitoylation, and recent studies suggest that these enzymes have a role in post-TGN trafficking (Wang *et al.*, 2003; Wang *et al.*, 2007) and the endocytic processing of the EGF receptors (Minogue *et al.*, 2006). Type III PI 4-kinases (α- and β-forms), on the other hand, are soluble enzymes structurally related to PI 3-kinases and

can be inhibited by higher concentrations of PI 3-kinase inhibitors, such as wortmannin (Wm). We have reported over 10 years ago that production of the agonist-sensitive phosphoinositide pools required the activity of Wm-sensitive type III PI 4-kinase enzymes (Nakanishi *et al.*, 1995). However, it is still not known which of the type III enzymes participates in the formation of the agonist-regulated PtdIns4*P* pools.

In the present study we used both pharmacological and genetic approaches to interfere with the activity of all of the PI 4-kinase enzymes and monitored phosphoinositide changes both by conventional metabolic labeling of inositol lipids and –phosphates and by employing GFP-tagged PH domains specifically recognizing PtdIns4*P* and PtdIns(4,5)P_2. These results show that the PH domain of the yeast OSH2 protein is a valuable tool to monitor plasma membrane PtdIns4*P* pools and that PI 4-kinase IIIα is required for sustained $InsP_3$ production and Ca^{2+} signaling. Although this conclusion was supported by siRNA-mediated gene silencing of the individual PI4K enzymes, the present studies also highlight the difficulties in obtaining conclusive data with siRNA on enzymes whose functions are near essential and emphasize the value of inhibitors to dissect the roles of these proteins in cellular signaling.

MATERIALS AND METHODS

Materials: Wortmannin and ionomycin were purchased from Calbiochem. Phenylarsine oxyde (PAO), β-mercaptoethanol, dithiothreitol and poly-Lysine were obtained from Sigma. Fura-2/AM and Pluronic acid were from Molecular Probes/Invitrogen. PIK93 was synthesized as previously described (Knight *et al.*, 2006). *Myo*-[^{3}H]inositol (60 Ci/mmol) was purchased from Amersham-Pharmacia and ortho-^{32}P-phosphate (9000

Ci/mmol) was from General Electric (NEN). The polyclonal antibody against PI4KIIIβ was purchased from UBI, the polyclonal antibody against PI4KIIα was a kind gift from Dr. Pietro De Camilli and the anti PI4KIIIα antibody was raised in New Zealand rabbits as previously described (Balla *et al.*, 2005).

DNA constructs and transfections: The OSH2-2x-GFP construct was generated by amplifying the PH domain sequence of OSH2 (residues: 256-424) from *S. cerevisiae* cDNA (ATCC) using two primer pairs to obtain fragments flanked by XhoI/EcoRI and EcoRI/KpnI sites. These fragments were then cloned in tandem between the XhoI/KpnI sites of the pEGFP-C1 plasmid (Clontech) with a linker (VNSKL) in between them following the design of Roy and Levine (Roy and Levine, 2004). The single PH domain version of the PH domain has also been created as well as the cyan and yellow fluorescent versions of the tandem construct. The PLCδ_1PH-GFP construct (Várnai and Balla, 1998) and its color variants have been previously described (Varnai *et al.*, 2002). The OSH1-PH-GFP was kindly provided by Dr. Mark Lemmon (Yu *et al.*, 2004), and the type IV phosphoinositide 5-phosphatase was a kind gift of Dr. Philip Majerus (Kisseleva *et al.*, 2000). The constructs used for the rapamycin-inducible translocation of the type-IV phosphoinositide 5-phosphatase have been described recently (Varnai *et al.*, 2006).

The HEK-293AT1 cells used for these studies have been stably transfected with the HA-AT1a and FLAG-AT1a angiotensin receptors in two rounds of selection using G418 and Zeocine for the two constructs, respectively. These cells were kindly provided by Drs. Alberto Jesus Olivares-Reyes and Kevin J. Catt. Cells were cultured in DMEM with Pen/Strep and 10% FBS and have been subjected to a G418/Zeo selection from time to time but not during cultures for the experiments.

For RNAi-mediated knock-down the cells were cultured either in 10 cm dishes (for suspension Ca^{2+} measurements), 12 well plates (for metabolic labeling studies) or 25 mm glass coverslips treated with poly-Lysine (for single cell Ca^{2+} or confocal studies). The duplexes used for treatment have been previously described (Balla *et al.*, 2005). Cells were treated with 100 nM siRNA twice in consecutive days and were analyzed on the 4th day after the first treatment.

Analysis of single cells for FRET, cytoplasmic Ca^{2+} measurements and confocal microscopy: HEK-293AT1 cells were cultured on glass coverslips ($3x10^5$ cells/35mm dish) pretreated with poly-Lysine and transfected with the various constructs (0.5-2μg DNA /dish) using Lipofectamine 2000 for 24 h as described elsewhere (Varnai *et al.*, 2005). For calcium measurements cells were loaded with 3 μM Fura-2/AM in Medium 199/Earle's balanced salt solution containing 1.2 mM $CaCl_2$, 3.6 mM KCl, 25 mM HEPES containing 1 mg/ml BSA, 0.06% Pluronic acid and 200 μM sulfinpyrazone, for 45 min at room temperature. Calcium measurements were performed at room temperature in a modified Krebs-Ringer buffer containing 120 mM NaCl, 4.7 mM KCl, 1.2 mM $CaCl_2$, 0.7 mM $MgSO_4$, 10 mM glucose, Na-Hepes 10 mM, pH 7.4. An Olympus IX70 inverted microscope equipped with a Lamda-DG4 illuminator and a MicroMAX-1024BFT digital camera and the appropriate filter sets was used for Ca^{2+} analysis. The same microscope equipped with a beam-splitter (Optical Insights) with a 505 nm dichroic mirror was used fro FRET analysis using 435/25 nm excitation and 475/30 And 535/30 emission filters, respectively. Images acquired simultaneously in the two emission wavelengths in the two halves of the camera sensor chip were used to form the ratios. These ratio values were then normalized taken their control initial values as 100 and the minimum values obtained after ionomycin (or agonist treatment if the latter was smaller)

as zero.

Cells expressing the OSH2-2x-PH-GFP and PLCd1PH-mRFP constructs were studied in the same modified Krebs-Ringer solution at 35 °C using a Zeiss 510Meta laser scanning confocal microscope (Zeiss, Thornwood, New York) in multi-track mode and pictures were taken in time-series mode. Membrane localization was assessed by forming membrane/cytosol intensity ratio values from line-intensity histograms of cells in the whole series of images post acquisition. Data acquisitions for Ca^{2+} and FRET measurements and processing were performed by the MetaFluor software (Molecular Devices). Post acquisition data analysis of the confocal images was performed with either the Metaview (Zeiss) or the Metamorph (Molecular Devices) software.

Cytoplasmic Ca^{2+} measurements in cells suspension: Cells grown on 10 cm culture plates were removed by mild trypsinization and loaded with 0.5 μM Fura-2/AM in the same solution described above for single Ca^{2+} measurements. Cells were then washed with the same medium without Fura-2/AM and stored at room temperature in the dark. Aliquots of cells (~ 5 x 10^5 cells) were centrifuged rapidly prior to the measurements and dispersed in 2.5 ml of the modified Krebs-Ringer buffer used for all other analysis. Ca^{2+}-measurements were performed at 35 C in a PTI Deltascan spectrofluorimeter.

Analysis of *myo*-[^{3}H]inositol- or ^{32}P-phopshate labeled lipids and [^{3}H]-labeled inositol phosphates: Cultured cells were labeled with *myo*-[^{3}H]inositol (20 μCi/ml) on 12-well culture plates in Ins-free DMEM supplemented with 2 % dialyzed FBS and 1 mg/ml BSA for 24 h. For ^{32}P-phopshate labeling, the cells were labeled with 2 μCi/ml *o*-^{32}P-phosphate for 3 hrs in phosphate-free DMEM supplemented with 1 mg/ml BSA. After *myo*-inositol labeling the cells were washed twice with a medium without inositol, and after a 10 min preincubation, inhibitors were added for a further 10 min before

stimulation with 100 nM AngII for the indicated times. Reactions were terminated by the addition of ice-cold perchloric acid (5% final concentration) and cells were kept on ice for 30 min. After scraping, and freezing/thawing, cells were centrifuged and the supernatant was processed for PCA extraction and analysis by high performance liquid chromatography as described previously (Nakanishi *et al.*, 1995). The cell pellet was also processed to extract the phosphoinositides by an acidic chloroform/methanol extraction followed by TLC analysis essentially as described previously (Nakanishi *et al.*, 1995). ^{32}P-labeled cells were not washed after the labeling period, but were treated in the same medium with inhibitors for 10 min followed by AngII (10^{-7} M) stimulation. Reactions were terminated by PCA and lipids were extracted and separated as with the inositol-labeled cells.

RESULTS

Pharmacological manipulations of type III PI 4-kinase activities affect agonist-regulated PtdIns4*P* pools.

High (μmolar) concentrations of Wm have been valuable tools to demonstrate a role of type III PI4Ks in $InsP_3/Ca^{2+}$ signaling. However, it is not possible to discriminate between the α and β forms of these PI4Ks based on their Wm sensitivities (Balla *et al.*, 1997; Knight *et al.*, 2006). Therefore, we took advantage of the recent characterization of isoform-specific PI 3-kinase inhibitors that revealed one of the inhibitors, PIK93, as being significantly more potent against PI4KIIIβ than PI4KIIIα (Knight *et al.*, 2006). This inhibitor inhibits PI 3-kinases and clearly discriminates between the two PI 4-kinase enzymes as assessed by an *in vitro* PI kinase assay on the purified mammalian proteins (Knight *et al.*, 2006). Unfortunately, none of the tested inhibitors showed the opposite

selectivity, i.e. inhibition of PI4KIIIα more potently than the PI4KIIIβ enzyme. Phenylarsine oxide (PAO) an SH reactive agent has been shown earlier to inhibit PI4Ks (Wiedemann *et al.*, 1996). PAO was found to inhibit type III PI 4-kinases whereas it is relatively ineffective against the type II enzymes based on *in vitro* kinase assays of the purified proteins (Balla *et al.*, 2002; Barylko *et al.*, 2002). Among the type III enzymes, PI4KIIIα is significantly more sensitive to PAO than the type IIIβ form (Balla *et al.*, 2002). This difference was also exploited to discriminate between the two enzymes in their involvement to generate the hormone-sensitive PtdIns4*P* pools. It is important to emphasize that PAO is a SH-reactive agent that probably has many targets in a cell. However, in the present study, the effects of PAO have only been tested on parameters that immediately reflect PI 4-kinase functions (see below) and therefore in this narrow context the effects of PAO on the purified PI 4-kinases and PtdIns4*P* formation in the intact cell can be correlated with somewhat higher confidence.

Several parameters were tested with these inhibitors in HEK293 cells stably expressing AT_{1a} angiotensin receptors (HEK-293-AT1). Cellular phosphoinositides were monitored after ^{32}P-phosphate or *myo*-[^{3}H]inositol labeling, and $InsP_3$ formation was followed from cells prelabeled with *myo*-[^{3}H]inositol. Finally, the effects of the inhibitors on AngII-induced changes in cytoplasmic Ca^{2+} concentration $[Ca^{2+}]_i$ were measured in cell suspensions with Fura-2. Fig. 1 shows the effects of inhibitors on $InsP_3$ levels analyzed from *myo*-[^{3}H]inositol-labeled cells and on Ca^{2+} signaling. As shown earlier in AngII stimulated adrenal glomerulosa cells (Nakanishi *et al.*, 1995), 10 μM Wm pretreatment greatly reduced the size of AngII induced $InsP_3$ elevation in HEK-293-AT1 cells and eliminated its sustained increase. This was also reflected in the cytoplasmic Ca^{2+} changes that became transient lacking the plateau phase of Ca^{2+} rise in Wm treated cells (Fig. 1B).

These effects of Wm were not observed at lower concentrations (below 300 nM) that already inhibit PI 3-kinases (data not shown), and were clearly caused by the limited supply of PtdIns4*P* and PtdIns(4,5)P_2 consistent with the inhibition of a PI4K that helps replenishing these pools during a sustained PLC activation. This is demonstrated in Fig. 2, where the ^{32}P-phosphate-labeled PtdIns4*P* and PtdIns(4,5)P_2 were analyzed in AngII stimulated cells. Without the inhibitors, AngII stimulation evokes a robust PLC activation resulting in the rapid depletion of PtdIns(4,5)P_2 that slowly returns toward prestimulatory levels. A similar change is observed in PtdIns4*P* levels due to the conversion of this lipid to PtdIns(4,5)P_2 by the PIP 5-kinases and its relatively slower synthesis by the PI4Ks. Pretreatment of the cells with 10 μM Wm greatly reduced the ^{32}P-labeled PtdIns4*P* and, hence, the AngII-induced changes observed in HEK-293-AT1 cells (Fig. 1A), but only slightly reduced basal PtdIns(4,5)P_2 levels. However, this treatment significantly enhanced the AngII-induced decrease in PtdIns(4,5)P_2 and strongly inhibited its resynthesis (Fig. 2B).

These effects of Wm were not reproduced by 250 nM PIK93, which inhibits PI4KIIIβ (and the PI 3-kinases) but not PI4KIIIα (Knight *et al.*, 2006), and which was found as effective as 10 μM Wm in inhibiting the ER to Golgi transport of ceramide, a process linked to PI4KIIIβ function (Toth *et al.*, 2006). PIK93 failed to affect either the $InsP_3$ and Ca^{2+} changes or those of the ^{32}P-labeled phosphoinositides during AngII-stimulation (Figs 1 and 2). The effect of PAO was tested at a concentration of 10 μM (applied in the presence of 1 mM β-mercaptoethanol to reduce its side effects). Although at this concentration PAO only partially inhibits PI4KIIIα (~80 %), it was chosen because it does not yet inhibit PI4KIIIβ (Balla *et al.*, 2002). As shown in Fig. 1, PAO partially mimicked the effects of Wm on both the $InsP_3$ and Ca^{2+} responses, substantially

reducing the sustained phases of both signals. Also, after 10 μM PAO treatment the ^{32}P-labeled phosphoinositides showed changes similar to those observed with Wm treatment (Fig. 2). All of these data were consistent with the limited supply of PtdIns4*P* in the presence of Wm or PAO but not PIK93 implicating the PI4KIIIα enzyme in the process, and the incomplete effects of PAO relative to Wm were consistent with the incomplete inhibition of PI4KIIIα at this concentration of the inhibitor.

Analysis of PtdIns(4,5)P_2 changes in the plasma membrane with the PLCδ$_1$PH domain.

These metabolic data provided information on the overall inositide pools of the cells but only limited information on the question of which membranes contributed to these changes. In fact, when cells were labeled with *myo*-[^{3}H]inositol, a smaller fraction of the inositides showed changes with AngII and the inhibitors (not shown) suggesting the presence of additional metabolic pools that become labeled with *myo*-[^{3}H]inositol in 24 h but not with ^{32}P-phosphate in 3 h. Therefore, further experiments were designed to monitor the phosphoinositide changes in single cells using pleckstrin homology (PH) domains that recognize PtdIns4*P* and PtdIns(4,5)P_2. PtdIns(4,5)P_2 changes were followed in single cells by either confocal microscopy or by FRET analysis using YFP and CFP-tagged versions of the PLCδ PH domain (van Der Wal *et al.*, 2001).

Cells were stimulated with AngII for the indicated times followed by the addition of high concentration (10 μM) of Ionomycin to activate PLC and to eliminate PLCδ$_1$PH-GFP localization and FRET (Várnai and Balla, 1998; Balla *et al.*, 2005). Subsequent addition of BAPTA (or EGTA) was used to reverse the Ca^{2+} change and allow the PtdIns(4,5)P_2 pools to recover. As shown in Fig. 3, the PLCδ$_1$PH translocation was rapid and complete after AngII treatment and the FRET signal rapidly returned close to the

baseline in control cells (black trace). Ionomycin treatment then caused the complete translocation of the probe to the cytosol from which the cells recovered slowly after Ca^{2+} chelation. These changes were not affected by preincubation of the cells with 250 nM PIK93 (Fig. 3, red trace) consistent with the previous data with metabolic labeling, suggesting that PI4KIIIβ plays no significant role in this process. In contrast, 10 μM PAO (with 1 mM β-mercaptoethanol) decreased the basal FRET by about 40% and completely prevented the relocalization of the probe after the AngII-induced complete translocation (Fig. 3, blue trace).

When the effects of Wm were tested in similar experiments, we encountered several technical difficulties. First, the inhibitory effects of Wm showed a strong light-induced reversal in single-cell experiments. At shorter excitation wavelengths these effects were stronger, making it essentially impossible to monitor the effect of Wm on single cell Ca^{2+} responses with Fura-2. (The photon flux density is significantly lower during Ca^{2+} measurements in cell-suspension, therefore those measurements are less sensitive to this effect). In addition, a significant drift was observed in the FRET baseline during the 10 min Wm preincubation period (without illumination). Although this was partially eliminated rapidly after 2-3 light scans, it was difficult to assess the basal FRET values after Wm treatment. Therefore the data on the effects of Wm (still partially inhibitory) were not included in this analysis.

Evaluation of the OSH2 PH domain as a probe of plasma membrane PtdIns4*P* levels.

None of the type III PI4K enzymes can be detected by immunofluorescence at the plasma membrane, yet PtdIns4*P* has to be present in the plasma membrane to be converted to PtdIns(4,5)P_2. To obtain information on the activity rather than the steady-

state cellular distribution of the PI4K enzymes, it was highly desirable to use reporter probes that monitor PtdIns4*P* production in single cells. PH domain-GFP chimeras with high *in vitro* PtdIns4*P* binding specificities, such as the FAPP1 or OSBP PH domains have been used for this purpose, but these probes mostly report on PtdIns4*P* formed in the Golgi as they also require Arf1-GTP for their Golgi recruitment (Levine and Munro, 2002; Godi *et al.*, 2004; Balla *et al.*, 2005). Although both of these PH domains can detect PtdIns4*P* formation in the plasma membrane under special circumstances (Balla *et al.*, 2005), they are not optimal for monitoring PtdIns4*P* in the plasma membrane. It has recently been shown by two independent studies that the PH domain of the yeast oxysterol binding protein homologue, OSH2 recognizes PtdIns4*P* in the plasma membrane, in spite of its limited specificity to bind PtdIns4*P* *in vitro* (Roy and Levine, 2004; Yu *et al.*, 2004). Therefore, first, we wanted to determine whether the OSH2 PH domain could, in fact, be used to follow PtdIns4*P* changes in the plasma membrane of mammalian cells.

Expression of the single PH domain of OSH2 fused to GFP showed clear plasma membrane localization and a strong nuclear staining as previously described (Roy and Levine, 2004; Yu *et al.*, 2004). Using two PH domains in tandem fused to GFP (GFP-OSH2-PH2x) greatly reduced the nuclear localization of the construct and clearly labeled the plasma membrane (Fig. 4A). Importantly, unlike in yeast cells, neither the tandem nor the single OSH2 PH domain decorated the Golgi membrane in COS-7 or HEK293 cells (Fig. 4A). In contrast, the yeast OSH1 PH domain labeled both the plasma membrane (at high expression levels) and the Golgi (Fig. 4A). Since the OSH2 PH domain has limited *in vitro* specificity to PtdIns4*P* (Yu *et al.*, 2004), we were concerned that it may be recruited to the plasma membrane by PtdIns(4,5)P_2. The specificity

question was examined with our recently developed method of acute depletion of PtdIns(4,5)P_2 in the plasma membrane. This approach is based on a drug induced plasma membrane recruitment of a truncated type-IV phosphoinositide 5-phosphatase (5-ptase) that was rendered cytoplasmic by mutations in its localization sequences (Varnai *et al.*, 2006). Recruitment of this cytoplasmic 5-ptase to the plasma membrane rapidly eliminated PtdIns(4,5)P_2 as monitored by PLCδ_1PH-GFP localization (Fig. 4B), without elimination of the GFP-OSH2-PH2x localization (Fig. 4C). This question was further analyzed in TIRF experiments where the release of the PH domain from the membrane can be better quantified. These experiments showed no effect of the phosphatase recruitment on GFP-OSH2-PH2x localization while eliminating PLCδ1PH-GFP localization in COS-7 cells (Korzeniowski and Balla, unpublished observations).

In a separate set of studies performed in COS-7 cells, the wild-type 5-ptase enzyme was expressed together with the mRFP-fused PLCδ1PH domain and the GFP-OSH2-PH2x construct. This triple transfection yielded many cells in which the plasma membrane localization of the PLCδ1PH-mRFP construct was eliminated indicating the depletion of PtdIns(4,5)P_2, yet the localization of the OSH2-PH2x was still preserved (Fig. 5A). These studies also confirmed that the OSH2-PH2x was not recruited to the membrane by PtdIns(4,5)P_2. When such cells were treated with 10 μM Wm, the localization of OSH2-PH2x was rapidly eliminated (Fig. 5A). Lower concentrations of Wm specific for PI 3-kinases had no such effect (not shown), indicating that the plasma membrane pool of PtdIns4*P* monitored by OSH2-PH2x requires the activity of type III PI 4-kinases. Notably, Wm exerted a much slower effect on OSH2-PH2x localization in cells not expressing the 5-phosphatase (Fig. 5B and see below) indicating that the

dephosphorylation of PtdIns(4,5)P_2 probably contributes to maintaining PtdIns4*P* levels in the membrane for a period of time when PI4K is inhibited.

The PH domain of OSH2 follows agonist-induced changes of PtdIns4*P* levels.

Next we determined whether GFP-OSH2-PH2x localization is affected by agonist-induced PLC activation. HEK-293-AT1 cells were co-transfected with the PLCδ1PH-mRFP and GFP-OSH2-PH2x for simultaneous monitoring of PtdIns(4,5)P_2 and PtdIns4*P*. As shown in Fig. 6, both constructs showed a rapid and transient translocation from the plasma membrane to the cytosol after AngII addition, but with notable kinetic differences. The translocation of the GFP-OSH2-PH2x occurred with a clearly measurable (about 5 seconds) delay compared to that of PLCδ1PH-mRFP. The kinetics of changes in GFP-OSH2-PH2x localization after AngII addition was indistinguishable regardless whether cells were preincubated with 10 mM Li^+ or not (not shown). Since Ins(1,4)P_2 levels accumulate to very high levels and remain elevated in Li^+-treated cells (e.g.(Balla *et al.*, 1988) due to the inhibition of the phosphatase that dephosphorylates Ins(1,4)P_2 (Majerus *et al.*, 1999), these findings suggest that the AngII-induced translocation of the GFP-OSH2-PH2x protein from the membrane is caused by the changes in membrane PtdIns4*P* levels rather than increases in Ins(1,4)P_2 levels in the cytosol. The slight delay in the response of GFP-OSH2-PH2x compared to that of PLCδ1PH-mRFP may reflect the slower (probably Ca^{2+} dependent) activation of the PLC isoform that hydrolyses PtdIns4*P*, or a slight delay in the conversion of PtdIns4*P* to PtdIns(4,5)P_2, but the contribution of the rapid Ins(1,4,5)P_3 increase to PLCδ1PH-mRFP translocation can also be responsible for its faster kinetics. Nevertheless, these data

together were consistent with the conclusion that the GFP-OSH2-PH2x reporter can monitor plasma membrane PtdIns4*P* changes during agonist activation.

Pharmacological manipulations of the membrane localization of OSH2-2x-PH.

In response to the PI 4-kinase inhibitors, the GFP-OSH2-PH2x domain showed changes in good agreement with the ^{32}P-phosphate and *myo*-[^{3}H]inositol labeling results: the localization of the GFP-OSH2-PH2x was monitored as a change in the membrane to cytosol ratio of fluorescence. (We also attempted to use FRET for analysis of OSH2-PH2x. However, the FRET signal was significantly smaller than with the PLCδ_1PH domain, which may reflect a lower abundance of PtdIns4*P* in the membrane or a lower density of the lipid in the membrane microdomains where it is found). The membrane-bound fluorescence of GFP-OSH2-PH2x already started to decrease during the 10 min Wm (10 μM) preincubation and about 70-80% of the membrane localization was eliminated by the end of this period. Lower concentrations of Wm (300 nM) did not show this effect (Fig. 7A). Subsequent addition of AngII rapidly eliminated the remaining localization and no re-localization of the probe was observed throughout the 10 min stimulation (Fig. 8A). It is important to note that the effects of Wm became almost undetectable when prolonged rapid sampling of the confocal pictures was used to follow localization of the PH domains. This was again attributed to the photo-lability of Wm inhibition discussed above and also observed in (Warashina, 1999). 10 μM PAO (with 1 mM β-mercaptoethanol) had a similar inhibitory effect on the basal localization, although this treatment did not evoke the same decrease as Wm did before AngII addition (Fig. 7B). The effect of PAO was completely reversed by simultaneous addition of 1 mM DTT (Fig. 7B). PAO also completely prevented the re-localization of the GFP-OSH2-

PH2x during AngII stimulation (Fig. 8B). In contrast, PIK93 (250 nM) had no effect on either the basal localization of the GFP-OSH2-PH2x probe (Fig. 7C) nor did it affect the response to AngII stimulation (Fig. 8A).

The effects of RNAi-mediated knock-down on phosphoinositides and Ca^{2+} signaling.

The results of the pharmacological analysis pointed to PI4KIIIα as the enzyme important in the maintenance of the agonist-sensitive phsophoinositide pools of the plasma membrane. We wanted to confirm these data with RNAi-mediated gene silencing. All PI4Ks were individually knocked down and the basal as well as AngII-induced changes in ^{32}P-labeled phosphoinositides and cytoplasmic Ca^{2+} were determined. These experiments showed that knock-down of either PI4Ks (confirmed in each experiments by Western blot analysis) (Fig. 9A) resulted in no appreciable change in the Ca^{2+} response of the cells in response to AngII regardless of whether these measurements were performed in cells suspension or in single attached cells (data not shown). ^{32}P-labeling of cells depleted in the respective PI4Ks showed variations due to the loss of cells in the PI4K down-regulated groups (Fig. 9B). To normalize for this difference, in each experiments the ratio of PtdIns4*P* and PtdIns(4,5)P_2 to the PtdIns/PtdA spots from unstimulated cells were calculated and compared between the control siRNA or PI4K siRNA treated groups. After such corrections, the labeling of both PtdIns4*P* and PtdIns(4,5)P_2 showed a ~25% decrease upon depletion of either of PI4KIIα, PI4KIIIα or PI4KIIIβ. This difference was statistically significant ($p<0.05$) for all enzyme knock-downs in PtdIns4*P* but only for PI4KIIα in PtdIns(4,5)P_2 (Fig. 9C). To assess PtdIns4*P* and PtdIns(4,5)P_2 synthesis during stimulation, the level of these lipids were measured after 10 min of AngII exposure, since by this time impaired resynthesis of these lipids was expected to be

detectable (see Fig.2 for full kinetics). In these experiments we also included treatment with 250 nM PIK93 before AngII treatment for each group to determine whether compensatory changes in PI4KIIIβ (although not apparent by Western analysis) could mask the effects of PI4KIIα or PI4KIIIα knock-down. Since there was absolutely no difference between the PIK93-AngII treated group and those only treated with AngII alone (not shown) these measurements have been combined for statistical analysis.

As shown in Fig. 9C, by 10 min of AngII treatment, PtdIns(4,5)P_2 and PtdIns4*P* levels returned to about 60-70% of their prestimulatory values (see also in Fig. 2) in control cells[1]. However, in cells depleted in PI4KIIIα, this value was significantly ($p<0.05$) lower (~50%) for both lipids, the difference being more pronounced in PtdIns(4,5)P_2 which could be measured with better accuracy. These differences were relatively minor and required a fine balance between siRNA treatment that still preserved enough cells for analysis yet showed sufficient knock-down to capture even these small changes.

In cells labeled with *myo*-[^{3}H]inositol the picture was further complicated as we observed an increased labeling of PtdIns4*P* and PtdIns(4,5)P_2 (and the inositol phosphates) in cells in which PI4KIIIα was depleted. This paradoxical change was attributed to a significant decrease in the free inositol level of PI4KIIIα knock-down cells and hence an increased specific activity of the labeled inositol pool (not shown). The reason for the consistent decrease in the level of inositol in cells depleted in PI4KIIIα is not known and is currently under further investigation.

DISCUSSION

In the present studies we asked the question of which of the PI4K enzymes is important for the generation of the plasma membrane pool of PtdIns4*P*, the precursor of PtdIns(4,5)P_2 during PLC activation in agonist-stimulated cells. This question has not been thoroughly addressed before for lack of appropriate tools to investigate it properly. In a previous study we showed that Wm-sensitive type III PI4Ks are required for the maintenance of agonist-sensitive phosphoinositide pools and hence Ins(1,4,5)P_3 and calcium signaling (Nakanishi *et al.*, 1995). These studies have been confirmed in other laboratories using different cell lines and different Ca^{2+}-mobilizing receptors and agonists (Willars *et al.*, 1998), but the identity of the enzyme(s) responsible for supplying PtdIns4*P* for the plasma membrane so far has eluded identification. The recent discovery of a PI 3-kinase inhibitor that discriminates between the type III PI4Ks has proven to be an invaluable tool to reinvestigate this question. Based on studies using ^{32}P-labeled phosphoinositides and *myo*-[^{3}H]inositol labeled inositol phosphates as well as cytoplasmic Ca^{2+} measurements, we were able to show that the effects of Wm are reproduced by inhibitors of PI4KIIIα but not PI4KIIIβ. It was also important to demonstrate that the PtdIns4*P* pool that is affected by the PI4KIIIα enzyme is in fact in the plasma membrane. This question was especially relevant, as none of the type III PI4Ks can be found in detectable amounts in the plasma membrane by immunfluorescence analysis of COS-7 or HEK-293 cells. Based on enzymatic characterization of the plasma membrane-associated PI4K activity performed two decades ago, it was concluded that the smaller and wortmannin-insensitive type II PI4Ks are present in the plasma membrane. This is supported by more recent immunocytochemical analysis that finds only the type II enzymes partially in the plasma membrane. Of the type III enzymes, PI4KIIIβ is found in the Golgi, and PI4KIIIα in the

ER/Golgi compartment [see (Balla and Balla, 2006) for citation of the original studies]. So it was of particular interest that the PH domain of the OSH2 protein, one of the yeast homologues of oxysterol binding proteins, is capable of reporting on changes in the plasma membrane that are consistent with changes of PtdIns4*P* levels.

These studies using PH domains of PLCδ_1 and OSH2-PH2x to monitor PtdIns(4,5)P_2 and PtdIns4*P*, respectively, confirmed the conclusions of the labeling and Ca^{2+} experiments that PI4KIIIα is the enzyme necessary for the maintenance of the plasma membrane pool of these lipids. The exact mechanism how this is achieved by the enzyme that is primarily ER/Golgi localized remains to be answered, but its is quite possible that a small fraction of the enzyme is in fact found at the plasma membrane regulated by a unique mechanism, while the larger fraction of the protein has additional functions in the ER/Golgi compartments. It is interesting to note that in *Saccharomyces cerevisiae* the Stt4p PI4K, a homologoue of PI4KIIIα, was reported to generate the plasma membrane PtdIns4*P* pool, but unlike in mammalian cells, the yeast enzyme is primarily found in the plasma membrane or a tightly associated compartment (Audhya and Emr, 2002). Paradoxically, there are well-documented ER and vacuolar functions of Stt4p in yeast, yet the protein has not been detected in those locations (Trotter *et al.*, 1998; Audhya *et al.*, 2000).

RNAi-mediated depletion of the individual PI4Ks has produced only small effects on ^{32}P-labeled PtdIns4*P* and PtdIns(4,5)P_2 but confirming the identity of the PI4KIIIα enzyme as one responsible for PtdIns4*P* and PtdIns(4,5)P_2 production at the plasma membrane during AngII stimulation. Although the levels of the enzymes can be knocked down to 15-30% of their original values, this level of knock-down produced only small effects in the labeled lipids and none on AngII-induced Ca^{2+} signaling. This probably

reflects the fact that small amounts of these enzymes are sufficient to support the production of this signaling pool of the lipids and cells with more severe knock-down could simply be eliminated or developed adaptive mechanisms to cope with the lack of the protein. This highlights the difficulties with knock-down studies of enzymes of crucial importance and the value of inhibitors that can be tested in short-term experiments.

In recent studies we were able to show the role of PI4Ks in other aspects of cellular PtdIns4*P* production where the knock-down data gave as clear results as the pharmacological approach. These included the recruitment of the FAPP1PH domain to the Golgi being mediated by both the PI4KIIIβ and PI4KIIα enzymes (Balla *et al.*, 2005), and the role of PI4KIIIβ in the ER to Golgi transport of ceramide (Toth *et al.*, 2006). In fact, in the former study we found that PI4KIIIα knock-down was able to decrease the plasma membrane recruitment of the FAPP1PH domain during recovery from a large Ca^{2+}-mediated PLC activation (Balla *et al.*, 2005). These studies collectively suggest that PI4Ks probably have multiple functions within the cells that are affected at different level of enzyme depletion.

Lastly, these studies utilizing the FAPP1PH, OSBP-PH and OSH2-PH domains highlight the values as well as the limitations of PH domain reporters. These studies clearly demonstrate that specific pools of PtdIns4*P* are detected by the individual probes and one reporter may not detect PtdIns4*P* in all locations within the cells. The FAPP1 and OSBP PH domains detect primarily the Golgi pool, although under special circumstances such as during recovery from massive Ca^{2+} induced PLC activation they can also detect PtdIns4*P* in the plasma membrane (Balla *et al.*, 2005). In contrast, the OSH2-2x-PH-GFP reporter detects PtdIns4*P* in the plasma membrane but not in the

Golgi compartment in mammalian cells. Only the yeast OSH1-PH decorates both the Golgi and the plasma membrane. Importantly, the plasma membrane localization of all of the PtdIns4*P* reporters depends on the activity of Wm-sensitive, type III PI4Ks, and based on the current studies specifically of the PI4KIIIα enzyme.

In summary, the present results establish PI4KIIIα as an important component of the generation of the plasma membrane pool of phosphoinositides. This conclusion is based on pharmacological studies and confirmed by RNAi-mediated gene silencing although the latter could not decrease the enzyme levels sufficiently to appreciably impair Ca^{2+} signaling. These studies also establish the OSH2-PH2x domain as a useful reporter to follow PtdIns4*P* changes in the plasma membrane. Further studies are in progress to identify the mechanism by which the apparently ER/Golgi localized PI4KIIIα can affect the supply of plasma membrane PtdIns4*P*.

ACKNOWLEDGEMENTS

We are grateful to Dr. Roger Y. Tsien for the monomeric red fluorescent protein, Dr Philip W. Majerus for the human type-IV 5-ptase clone and Drs. Alberto-Jesus Olivares-Reyes and Kevin J. Catt for the HEK-293-AT1 cells. The confocal imaging was performed at the Microscopy & Imaging Core of the National Institute of Child Health and Human Development, NIH with the kind assistance of Drs. Vincent Schram and James T. Russell. This research was supported in part by the Intramural Research Program of the National Institute of Child Health and Human Development of the National Institutes of Health, and by an appointment of PV to the Senior Fellowship

Program at the NIH. This latter program is administered by the Oak Ridge Institute for Science and Education through an interagency agreement between the U.S. Department of Energy and the National Institutes of Health. A. B and P.V. are also Bolyai Fellows of the Hungarian Academy of Science and were supported by the Hungarian Scientific Research fund (OTKA NF-68563 and PF 63893) and the Medical Research Council (ETT 440/2006).

ABBREVIATIONS:

AngII, angiotensin II; 5-ptase, type-IV phosphoinositide 5-phosphatase; DTT, dithiothreitol; ER, endoplasmic reticulum; FAPP, four phosphate adaptor protein; FRET, fluorescence resonance energy transfer; GFP, enhanced green fluorescent protein; $InsP_3$, inositol trisphosphate; Mer, β-mercaptoethanol; mRFP, monomeric red fluorescent protein; OSBP, oxysterol binding protein; PAO, Phenylarsine oxide; PI4K, phosphatidylinositol 4-kinase; PtdIns4*P*, phosphatidylinositol 4-phosphate; PtdIns(4,5)P_2, phosphatidylinositol 4,5-bisphosphate; PtdA, phosphatidic acid; PLC, phospholipase C; PH, pleckstrin homology; siRNA, small interfering RNA; Wm, wortmannin

FOOTNOTES:

[1] Since AngII treatment increases the labeling of PtdIns/PtdA, and the knock-down of the individual enzymes could also affect this response, the correction for cell loss due to knock-down of the enzymes was based on the PtdIns/PtdA values obtained in the unstimulated cells also for the in AngII-stimulated samples. We assumed that AngII treatment does not cause cells loss.

REFRENCES

Audhya, A., and Emr, S.D. (2002). Stt4 PI 4-kinase localizes to the plasma membrane and functions in the Pkc1-mediated MAP kinase cascade. Dev.Cell *2*, 593-605.

Audhya, A., Foti, M., and Emr, S.D. (2000). Distinct roles for the yeast phosphatidylinositol 4-kinases, stt4p and pik1p, in secretion, cell growth, and organelle membrane dynamics. Mol.Biol.Cell *11*, 2673-2689.

Balla, A., and Balla, T. (2006). Phosphatidylinositol 4-kinases; old enzymes with emerging functions. Trends Cell Biol *16*, 351-361.

Balla, A., Tuymetova, G., Barshishat, M., Geiszt, M., and Balla, T. (2002). Characterization of type II phosphatidylinositol 4-kinase isoforms reveals association of the enzymes with endosomal vesicular compartments. J.Biol.Chem. *277*, 20041-22050.

Balla, A., Tuymetova, G., Tsiomenko, A., Varnai, P., and Balla, T. (2005). A plasma membrane pool of phosphatidylinositol 4-phosphate is generated by phosphatidylinositol 4-kinase type-III alpha: studies with the PH domains of the oxysterol binding protein and FAPP1. Mol.Biol.Cell *16*, 1282-1295.

Balla, T., Baukal, A.J., Guillemette, G., and Catt, K.J. (1988). Multiple pathways of inositol polyphosphate metabolism in angiotensin-stimulated adrenal glomerulosa cells. J.Biol.Chem. *263*, 4083-4091.

Balla, T., Downing, G.J., Jaffe, H., Kim, S., Zolyomi, A., and Catt, K.J. (1997). Isolation and molecular cloning of wortmannin-sensitive bovine type-III phosphatidylinositol 4-kinases. J.Biol.Chem. *272*, 18358-18366.

Barylko, B., Wlodarski, P., Binns, D.D., Gerber, S.H., Earnest, S., Sudhof, T.C., Grichine, N., and Albanesi, J.P. (2002). Analysis of the catalytic domain of phosphatidylinositol 4-kinase type II. J Biol Chem *277*, 44366-44375.

Berridge, M.J., and Irvine, R.F. (1984). Inositol trisphosphate, a novel second messenger in cellular signal transduction. Nature *312*, 315-321.

Creba, J.A., Downes, C.P., Hawkins, P.T., Brewster, G., Michell, R.H., and Kirk, C.J. (1983). Rapid breakdown of phosphatidylinositol 4-phosphate and phosphatidylinositol 4,5-bisphosphate in rat hepatocytes stimulated by vasopressin and other Ca2+-mobilizing hormones. Biochem.J. *212*, 733-747.

Doughman, R.L., Firestone, A.J., and Anderson, R.A. (2003). Phosphatidylinositol phosphate kinases put PI4,5P(2) in its place. J Membr Biol *194*, 77-89.

Godi, A., Di Campi, A., Konstantakopoulos, A., Di Tullio, G., Alessi, D.R., Kular, G.S., Daniele, T., Marra, P., Lucocq, J.M., and De Matteis, M.A. (2004). FAPPs control Golgi-to-cell-surface membrane traffic by binding to ARF and PtdIns(4)P. Nat.Cell Biol. *6*, 393-404.

Kisseleva, M.V., Wilson, M.P., and Majerus, P.W. (2000). The isolation and characterization of a cDNA encoding phospholipid-specific inositol polyphosphate 5-phosphatase. J.Biol.Chem. *275*, 20110-20116.

Knight, Z.A., Gonzalez, B., Feldman, M.E., Zunder, E.R., Goldenberg, D.D., Williams, O., Loewith, R., Stokoe, D., Balla, A., Toth, B., Balla, T., Weiss, W.A., Williams, R.L., and Shokat, K.M. (2006). A Pharmacological Map of the PI3-K Family Defines a Role for p110alpha in Insulin Signaling. Cell *125*, 733-747.

Levine, T.P., and Munro, S. (2002). Targeting of Golgi-specific pleckstrin homology domains involves both PtdIns 4-kinase-dependent and -independent components. Curr.Biol. *12*, 695-704.

Majerus, P.W., Kisseleva, M.V., and Norris, F.A. (1999). The role of phosphatases in inositol signaling reactions. J.Biol.Chem. *274*, 10669-10672.

Minogue, S., Waugh, M.G., De Matteis, M.A., Stephens, D.J., Berditchevski, F., and Hsuan, J.J. (2006). Phosphatidylinositol 4-kinase is required for endosomal trafficking and degradation of the EGF receptor. J Cell Sci *119*, 571-581.

Nakanishi, S., Catt, K.J., and Balla, T. (1995). A wortmannin-sensitive phosphatidylinositol 4-kinase that regulates hormone-sensitive pools of inositolphospholipids. Proc.Natl.Acad.Sci.U.S.A. *92*, 5317-5321.

Roy, A., and Levine, T.P. (2004). Multiple pools of phosphatidylinositol 4-phosphate detected using the pleckstrin homology domain of Osh2p. J.Biol.Chem. *279*, 44683-44689.

Toth, B., Balla, A., Ma, H., Knight, Z.A., Shokat, K.M., and Balla, T. (2006). Phosphatidylinositol 4-kinase IIIbeta regulates the transport of ceramide between the endoplasmic reticulum and Golgi. J Biol Chem *281*, 36369-36377.

Trotter, P.J., Wu, W.I., Pedretti, J., Yates, R., and Voelker, D.R. (1998). A genetic screen for aminophospholipid transport mutants identifies the phosphatidylinositol 4-kinase, Stt4p, as an essential component in phosphatidylserine metabolism. J.Biol.Chem. *273*, 13189-13196.

van Der Wal, J., Habets, R., Varnai, P., Balla, T., and Jalink, K. (2001). Monitoring Phospholipase C activation kinetics in live cells by FRET. J.Biol.Chem. *276*, 15337-15344.

Varnai, P., Balla, A., Hunyady, L., and Balla, T. (2005). Targeted expression of the inositol 1,4,5-triphosphate receptor (IP3R) ligand-binding domain releases Ca2+ via endogenous IP3R channels. Proc Natl Acad Sci U S A *102*, 7859-7864.

Várnai, P., and Balla, T. (1998). Visualization of phosphoinositides that bind pleckstrin homology domains: calcium-and agonist-induced dynamic changes and relationship to myo-[3H]inositol-labeled phosphoinositide pools. J.Cell Biol. *143*, 501-510.

Varnai, P., Lin, X., Lee, S.B., Tuymetova, G., Bondeva, T., Spat, A., Rhee, S.G., Hajnoczky, G., and Balla, T. (2002). Inositol lipid binding and membrane localization of isolated pleckstrin homology (PH) domains. Studies on the PH domains of phospholipase C delta 1 and p130. J.Biol.Chem. *277*, 27412-27422.

Varnai, P., Thyagarajan, B., Rohacs, T., and Balla, T. (2006). Rapidly inducible changes in phosphatidylinositol 4,5-bisphosphate levels influence multiple regulatory functions of the lipid in intact living cells. J Cell Biol *175*, 377-382.

Wang, J., Sun, H.Q., Macia, E., Kirchhausen, T., Watson, H., Bonifacino, J.S., and Yin, H.L. (2007). PI4P Promotes the Recruitment of the GGA Adaptor Proteins to the Trans-Golgi Network and Regulates Their Recognition of the Ubiquitin Sorting Signal. Mol Biol Cell.

Wang, Y.J., Li, W.H., Wang, J., Xu, K., Dong, P., Luo, X., and Yin, H.L. (2004). Critical role of PIP5KIgamma 87 in InsP3-mediated Ca(2+) signaling. J.Cell Biol. *167*, 1005-1010.

Wang, Y.J., Wang, J., Sun, H.Q., Martinez, M., Sun, Y.X., Macia, E., Kirschhausen, T., Albanesi, J.P., Roth, M.G., and Yin, H.L. (2003). Phosphatidylinositol 4 phosphate regulates targeting of clathrin adaptor AP-1 complexes to the Golgi. Cell *114*, 299-310.

Warashina, A. (1999). Light-evoked recovery from wortmannin-induced inhibition of catecholamine secretion and synaptic transmission. Arch.Biochem.Biophys. *367*, 303-310.

Wiedemann, C., Schäfer, T., and Burger, M.M. (1996). Chromaffin granule-associated phosphatidylinositol 4-kinase activity is required for stimulated secretion. EMBO J. *15*, 2094-2101.

Willars, G.B., Nahorski, S.R., and Challiss, R.A. (1998). Differential regulation of muscarinic acetylcholine receptor-sensitive polyphosphoinositide pools and consequences for signaling in human neuroblastoma cells. J Biol Chem *273*, 5037-5046.

Yu, J.W., Mendrola, J.M., Audhya, A., Singh, S., Keleti, D., DeWald, D.B., Murray, D., Emr, S.D., and Lemmon, M.A. (2004). Genome-wide analysis of membrane targeting by S. cerevisiae pleckstrin homology domains. Mol.Cell *13*, 677-688.

LEGEND TO FIGURES

Figure 1. Effects of PI4K inhibitors on the kinetics of $InsP_3$ and $[Ca^{2+}]_i$ changes in HEK-293-AT1 cells stimulated with AngII. (A) HEK-293-AT1 cells were labeled with *myo*-[^{3}H]inositol for 24 h in inositol-free medium as described under the Materials and Methods. After washing, AngII (10^{-7} M) was added to the cells for the indicated times and reactions terminated by PCA. Labeled inositol phosphates were extracted from the soluble fraction and separated by HPLC connected to a scintillation flow detector as previously described (Nakanishi *et al.*, 1995). Data are shown are means ± range of duplicate determinations. The concentrations of the inhibitors added 10 min prior to AngII were: 250 nM PIK93, 10 μM PAO with 1 mM β-mercaptoethanol (Mer), and 10 μM wortmannin (Wm). Two additional experiments were performed with the 10 min time points with similar results. (B) HEK-293-AT1 cells were loaded with Fura-2/AM and their fluorescence monitored in a fluorescent spectrophotometer as described under Materials and Methods. AngII (10^{-7} M) was added at the indicated time. Pretreatment with the inhibitors for 10 min was as described in panel A. This result is a representative of three similar observations.

Figure 2. Effects of PI4K inhibitors on the kinetics of phosphoinositide changes in HEK-293-AT1 cells stimulated with AngII. HEK-293-AT1 cells were labeled with ^{32}P-phosphate for 3 h in a phosphate-free medium as described under the Materials and

Methods. AngII (10^{-7} M) was added to the cells at the indicated times and reactions terminated by the addition of PCA. Lipids were extracted from the cell pellets, separated by TLC and analyzed both by a phosphorimager and scintillation counting of the spots cut out from the plates. Data shown are means ± range of duplicate determinations. The concentrations of the inhibitors added 10 min prior to AngII were: 250 nM PIK93, 10 μM PAO with 1 mM β-mercaptoethanol (Mer), and 10 μM wortmannin (Wm). This full time course experiment was repeated from *myo*-[^{3}H]inositol labeled cells with similar results supporting the same conclusion.

Figure 3. Effects of PI4K inhibitors on the kinetics of redistribution of PLCδ_1PH domain in HEK-293-AT1 cells stimulated with AngII. HEK-293 cells stably transfected with the AT1a angiotensin receptors were co-transfected with the PLCd1PH-CFP and –YFP constructs for 24 h. The FRET signal from individual cells was then monitored in a wide-filed fluorescent microscope equipped with an emission beam splitter using 430 nm excitation and 535 and 475 emissions as described under Materials and Methods. To minimize light exposure, after the 1st min of stimulation, data were collected in longer intervals. AngII (10^{-7} M) was added at the indicated time followed by 10 μM inonomycin (Iono) and 5 mM BAPTA (or EGTA). The dark bar represents the exposure of the cells to the high (extracellular) Ca^{2+} concentration following ionomycin treatment. The concentrations of the inhibitors added 10 min prior to AngII were: 250 nM PIK93, 10 μM PAO with 1 mM β-mercaptoethanol (Mer). FRET was expressed as fluorescent emission ratio values (535/475 nm) and was normalized so that the ratio value recorded before the addition of inhibitors was taken as 100% and the lowest values (after

AngII or ionomycin) were taken as zero percent. Means ± S.E.M. (or range) from 5, 2 and 3 cells are shown for control, PAO+Mer and PIK93, respectively. Similar changes were observed in two additional experiments.

Figure 4. Localization of OSH1- and OSH2-PH domain-GFP fusion proteins in HEK-293-AT1 cells. (A) The PH domains of the yeast oxysterol binding protein homologues, OSH2 and OSH1 were fused to GFP as described under Materials and Methods. These constructs were transfected into HEK-293-AT1 cells and examined 24 h post transfection with live cell confocal microscopy. Note that the OSH2-PH domain binds to the plasma membrane and strongly accumulates in the nucleus but does not bind to the Golgi. A tandem PH domain of the OSH2 protein shows smaller signal in the nucleus and a strong plasma membrane binding. The OSH1-PH domain binds both the plasma membrane and the Golgi. (B) Elimination of the plasma membrane localization of $PLC\delta_1PH$-GFP (monitoring $PtdIns(4,5)P_2$) by plasma membrane recruitment of a phosphoinositide 5-phosphatase. HEK-293-AT1 cells were transfected with a truncated type-IV phosphoinositide 5-phosphatase fused to mRFP and FKBP12, a plasma membrane targeted FRB-CFP construct and the $PLC\delta_1PH$-YFP reporter described in (Varnai *et al.*, 2006). Addition of rapamycin for 3 min recruits the otherwise cytoplasmic 5-ptase construct to the plasma membrane (left panels) with a concomitant elimination of $PtdIns(4,5)P_2$ and loss of $PLC\delta_1PH$-YFP localization (middle panels). (C) The same manipulations do not eliminate the plasma membrane localization of the OSH2-PH2x-GFP suggesting that this construct is not kept at the membrane by $PtdIns(4,5)P_2$.

Figure 5. Localization of OSH2-PH2x-GFP to the plasma membrane is wortmannin-sensitive. (A) COS-7 cells were transfected with OSH2-PH2x-GFP together with PLCδ_1PH-mRFP and the wild-type type IV phosphoinositide 5-phosphatase for 24 h. Cells were selected so that the PLCδ_1PH-mRFP showed no localization indicating the lack of PtdIns(4,5)P_2 as a result of phosphatase expression. These cells still showed plasma membrane localization of OSH2-PH2x-GFP indicating that the construct is kept in the membrane not by PtdIns(4,5)P_2. Addition of 10 μM wortmannin (Wm) to such cells caused a rapid translocation of the OSH2-PH2x-GFP domain construct from the membrane to the cytosol. (B) Release of the OSH2-PH2x-GFP construct from the membrane after Wm treatment is significantly slower in control cells where PtdIns(4,5)P_2 is present in the membrane.

Figure 6. Angiotensin II-induced changes in membrane phosphoinositides simultaneously monitored by OSH2-PH2x-GFP and PLCδ_1PH-mRFP. HEK-293 cells stably transfected with the AT1a angiotensin receptors were co-transfected with the PLCd1PH-mRFP and OSH2-PH2x-GFP constructs for 24 h. Live cells were examined in a confocal microscope at 35 ^{o}C during stimulation with AngII (10^{-7} M) and pictures were taken every 5 seconds. Note the faster response observed with the PLCδ_1PH-mRFP. Quantification of membrane/cytosolic fluorescence intensities as function of time calculated from line-intensity histograms from 5-7 cells obtained in two separate experiments (Means ± S.E.M)

Figure 7. Effects of PI4K inhibitors on the membrane localization of the OSH2-PH2x-GFP. HEK-293 cells stably transfected with the AT1a angiotensin receptors were

co-transfected with the OSH2-PH2x-GFP constructs for 24 h. Live cells were examined in a confocal microscope at 35 °C. Calculation of membrane/cytosolic fluorescence intensities as function of time was performed from line-intensity histograms from images taken at every minute during a10 min incubation with the inhibitors. The concentrations of the inhibitors were: 250 nM PIK93, 10 μM PAO with 1 mM β-mercaptoethanol (Mer), 10 μM PAO with 1 mM dithiotreitol (DTT) and 10 μM or 300 nM wortmannin (Wm). Means ± S.E.M. from 3-25 cells from 2-4 separate experiments are shown.

Figure 8. Effects of AngII on the membrane localization of the OSH2-PH2x-GFP after treatment with PI4K inhibitors. HEK-293 cells stably transfected with the AT1a angiotensin receptors were co-transfected with the OSH2-PH2x-GFP constructs for 24 h. Live cells were examined in a confocal microscope at 35 °C. Calculation of membrane/cytosolic fluorescence intensities as function of time was performed from line-intensity histograms from images taken at the indicated times after AngII addition. The concentrations of the inhibitors applied for 10 min prior AngII were: 250 nM PIK93, 10 μM PAO with 1 mM β-mercaptoethanol (Mer), and 10 μM wortmannin (Wm). Means ± S.E.M. from 3-25 Cells from 2-4 separate experiments are shown. The initial ratio values of Wm treated cells are higher in these experiments than those shown in Fig. 7 at the end of Wm treatment, as cells with still measurable OSH2-PH2x-GFP localization after Wm treatment were selected for this analysis.

Figure 9. Effects of knock-down of the individual PI 4-kinases on ^{32}P-phosphate labeling of phosphoinositides and their response to AngII stimulation in HEK-293-AT1 cells. HEK-293 cells stably transfected with the AT1a receptors were treated with

siRNA for 3 days as described under Materials and Methods. The extent of knock-down was determined by Western blot analysis (A). On the 4th day, cells were labeled with ^{32}P-phosphate for 3 h in phosphate-free medium. Cells were then either treated with AngII (10^{-7} M) or saline, and incubated for an additional 10 min. Reactions were terminated by PCA and lipids were extracted from the cell pellets separated by TLC and analyzed by a PhosphorImager (B). Due to cell loss in the PI4K depleted cells, the individual PtdIns4*P* and PtdINs(4,5)P_2 spots were normalized to the PtdA/PtdIns values measured in unstimulated samples of the respective groups. These ratios were then compared between the PI4K down-regulated or control siRNA-treated cells in each of the three experiments performed in duplicates (C). In each experiments, the effect of 250 nM PIK93 was also determined in the AngII group. Since PIK93 treated cells showed no difference compared to those treated with AngII only, these values were pooled in the statistical analysis. Means ± S.E.M are shown and the p values obtained in one sample t-tests as compared to control siRNA-treated cells are shown above the bars.

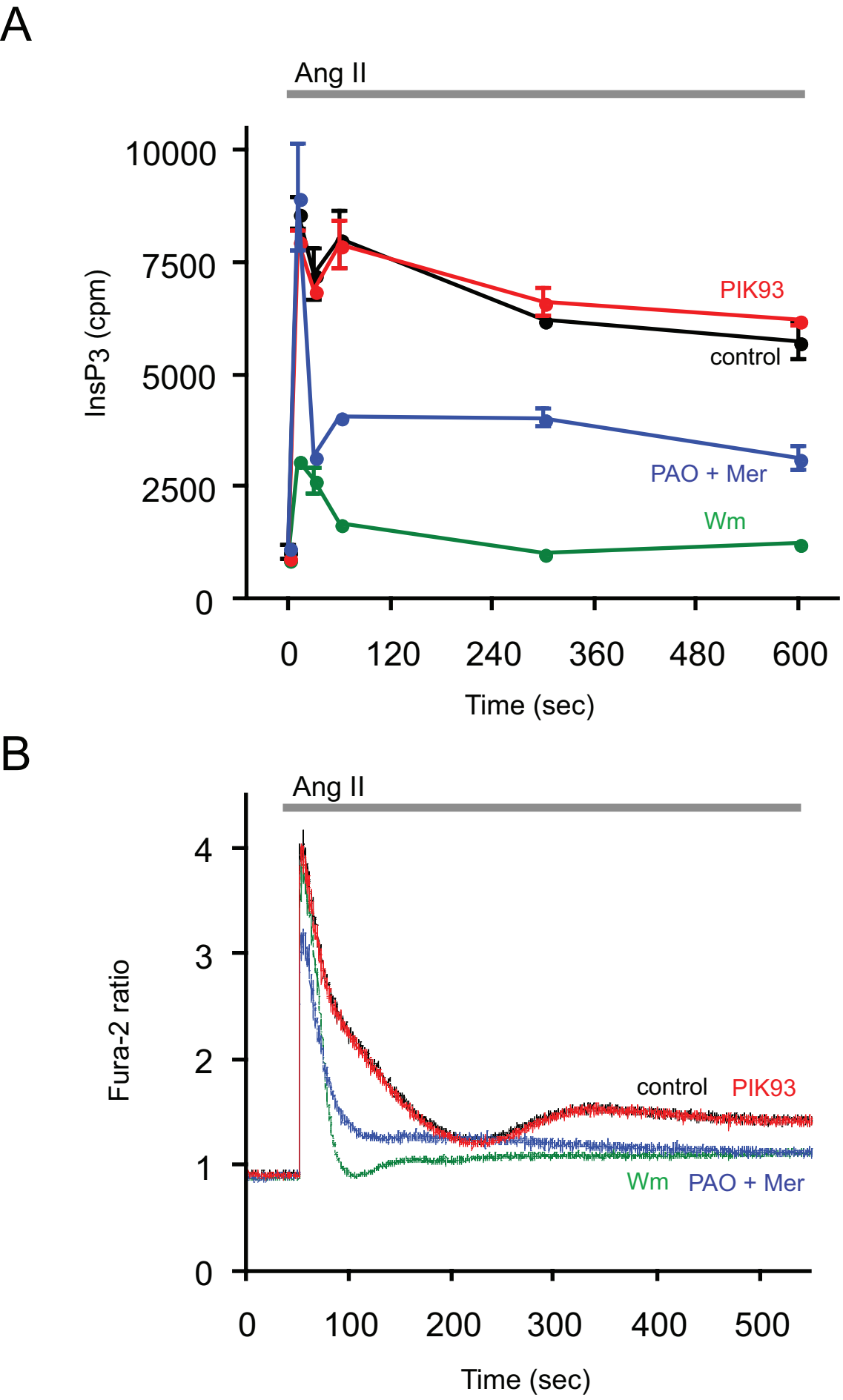
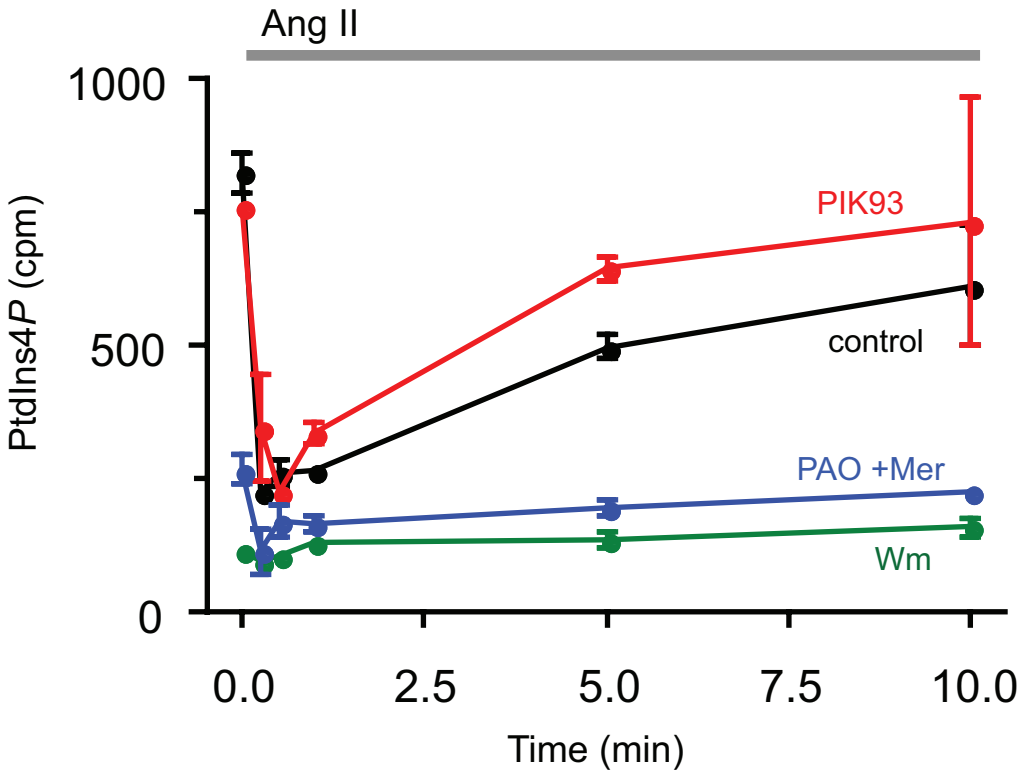

A
Ang II
10000
7500
5000
2500
0
InsP3 (cpm)
PIK93
control
PAO + Mer
Wm
0
120
240
360
480
600
Time (sec)
B
Ang II
4
3
2
1
0
Fura-2 ratio
control
PIK93
Wm
PAO + Mer
0
100
200
300
400
500
Time (sec)

Figure 1

A


Ang II
1000
500
0
PtdIns4P (cpm)
PIK93
control
PAO +Mer
Wm
0.0
2.5
5.0
7.5
10.0
Time (min)

B

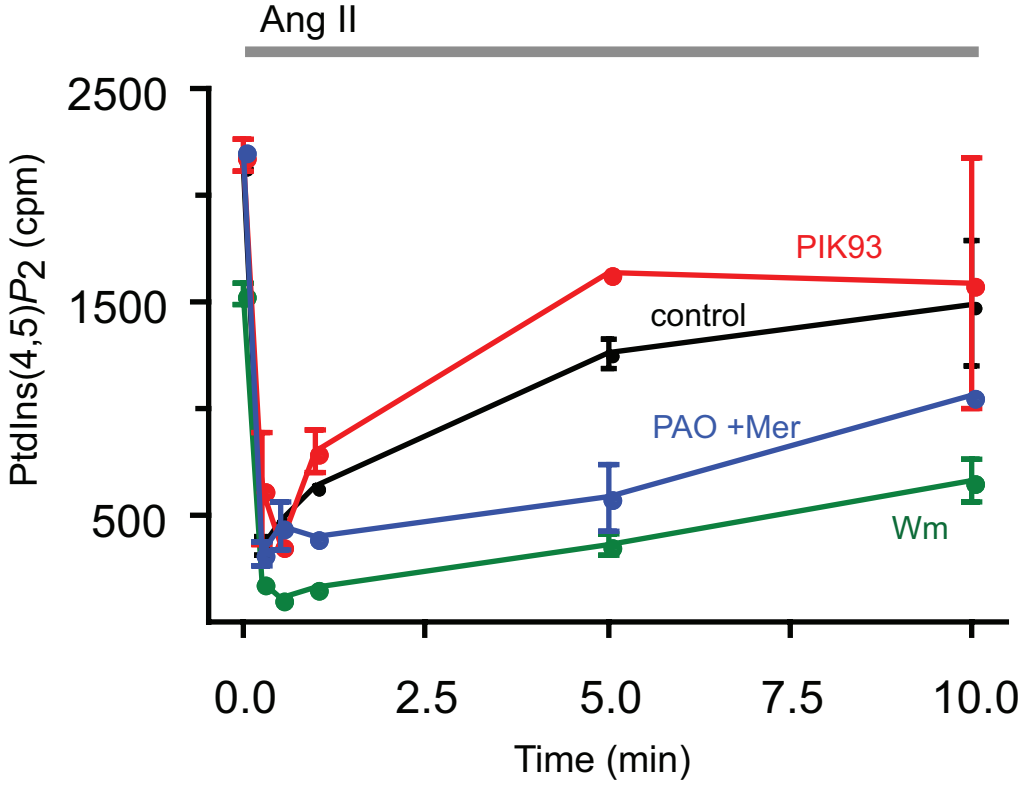

Ang II
2500
1500
500
PtdIns(4,5)P2 (cpm)
PIK93
control
PAO +Mer
Wm
0.0
2.5
5.0
7.5
10.0
Time (min)

Figure 2

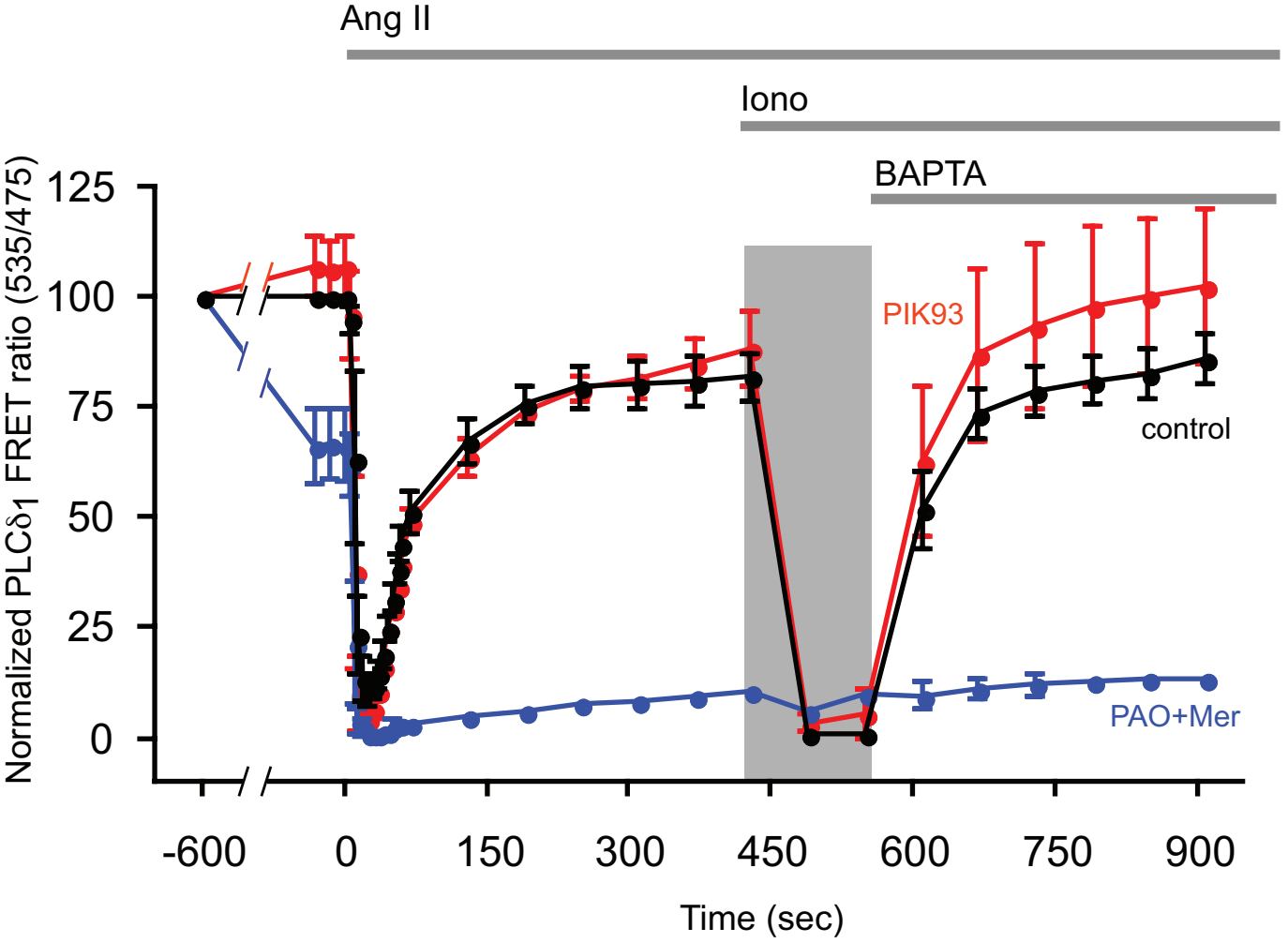

Ang II
Iono
BAPTA
PIK93
control
PAO+Mer
Normalized PLCδ1 FRET ratio (535/475)
125
100
75
50
25
0
-600
0
150
300
450
600
750
900
Time (sec)

Figure 3

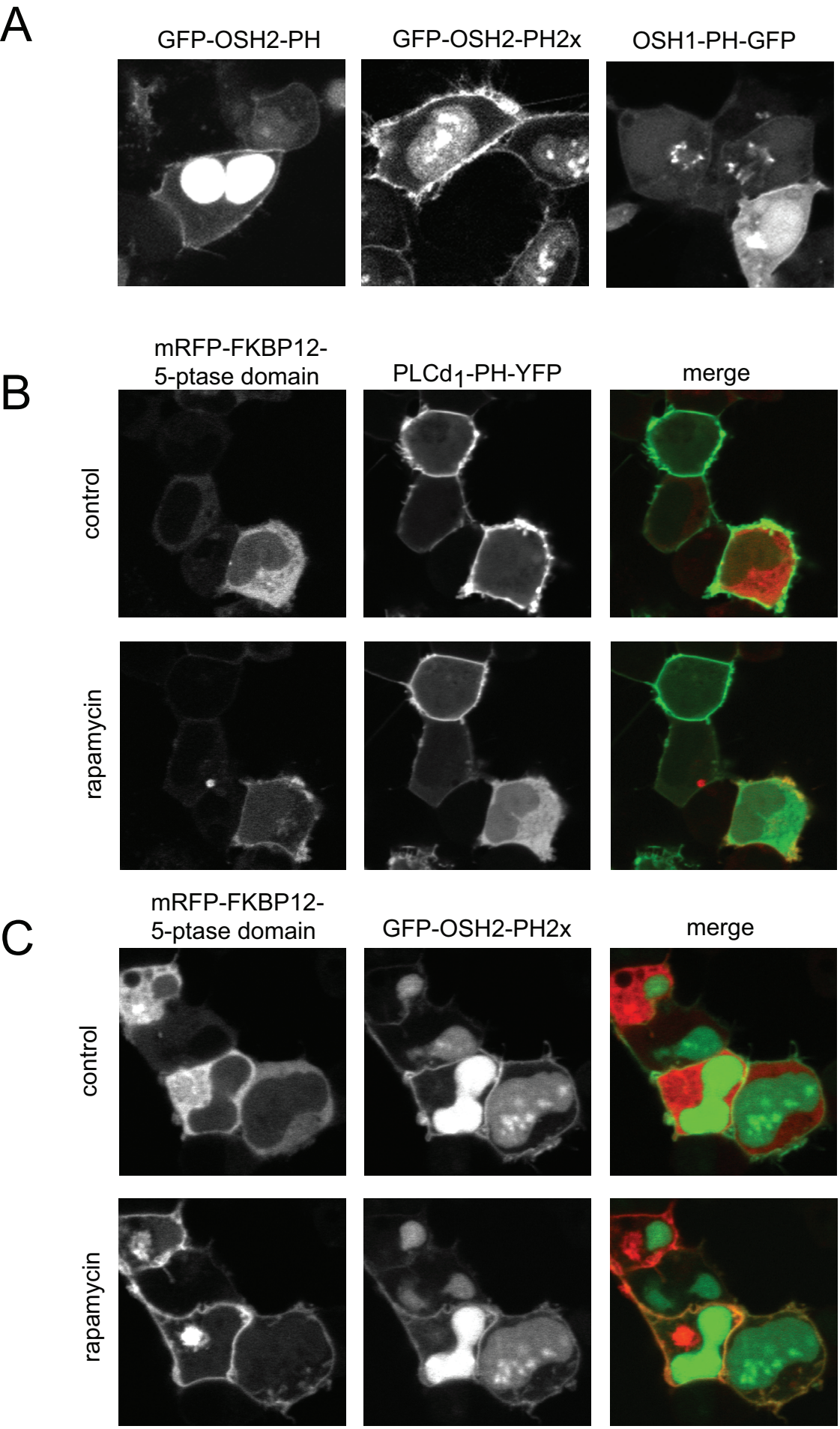
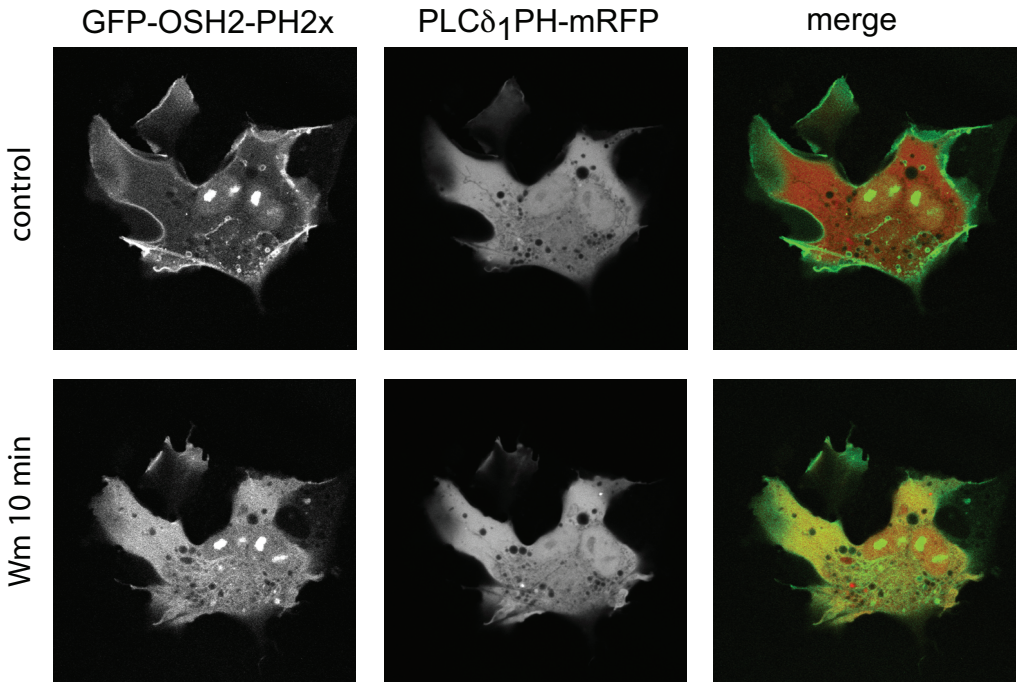

A
GFP-OSH2-PH
GFP-OSH2-PH2x
OSH1-PH-GFP
B
mRFP-FKBP12-
5-ptase domain
PLCd1-PH-YFP
merge
control
rapamycin
C
mRFP-FKBP12-
5-ptase domain
GFP-OSH2-PH2x
merge
control
rapamycin

Figure 4

A


GFP-OSH2-PH2x
PLCδ1PH-mRFP
merge
control
Wm 10 min

B

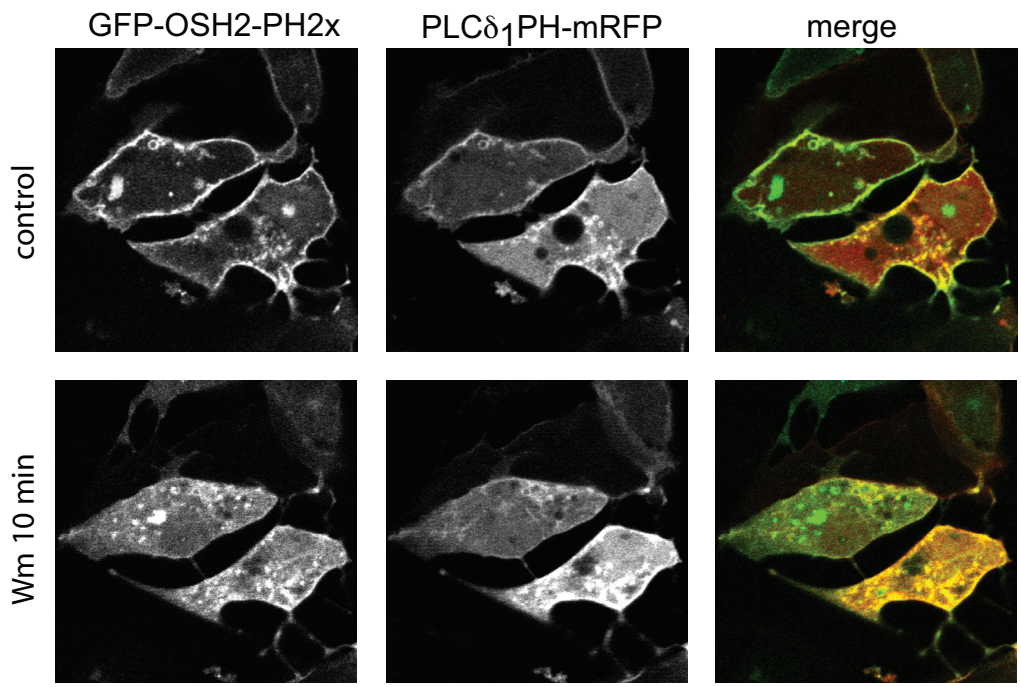
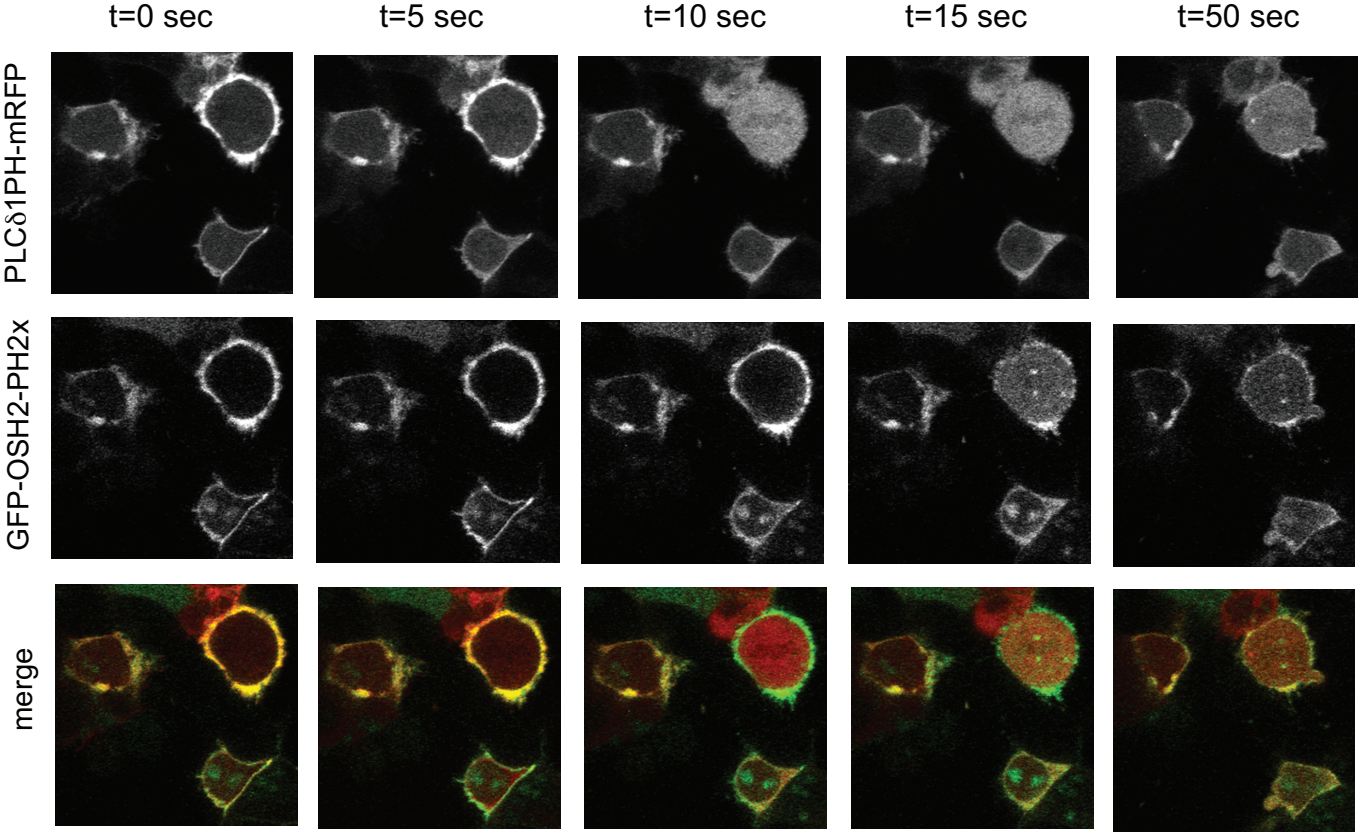

GFP-OSH2-PH2x
PLCδ1PH-mRFP
merge
control
Wm 10 min

Figure 5


t=0 sec
t=5 sec
t=10 sec
t=15 sec
t=50 sec
PLCδ1PH-mRFP
GFP-OSH2-PH2x
merge

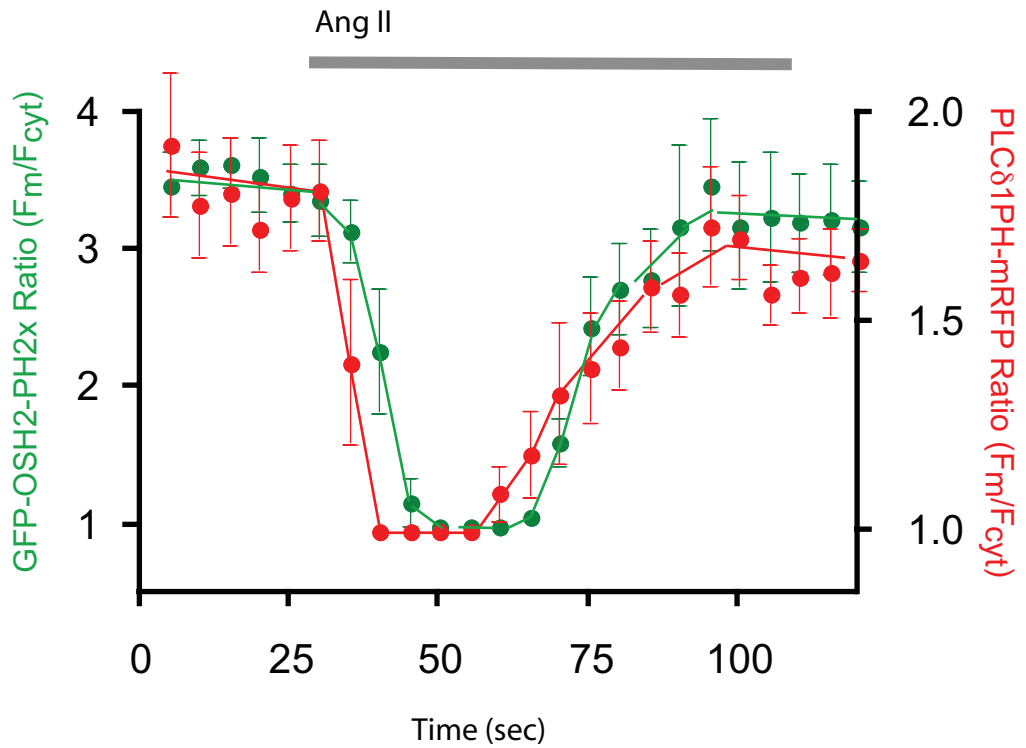

Ang II
GFP-OSH2-PH2x Ratio (Fm/Fcyt)
PLCδ1PH-mRFP Ratio (Fm/Fcyt)
4
3
2
1
2.0
1.5
1.0
0
25
50
75
100
Time (sec)

Figure 6

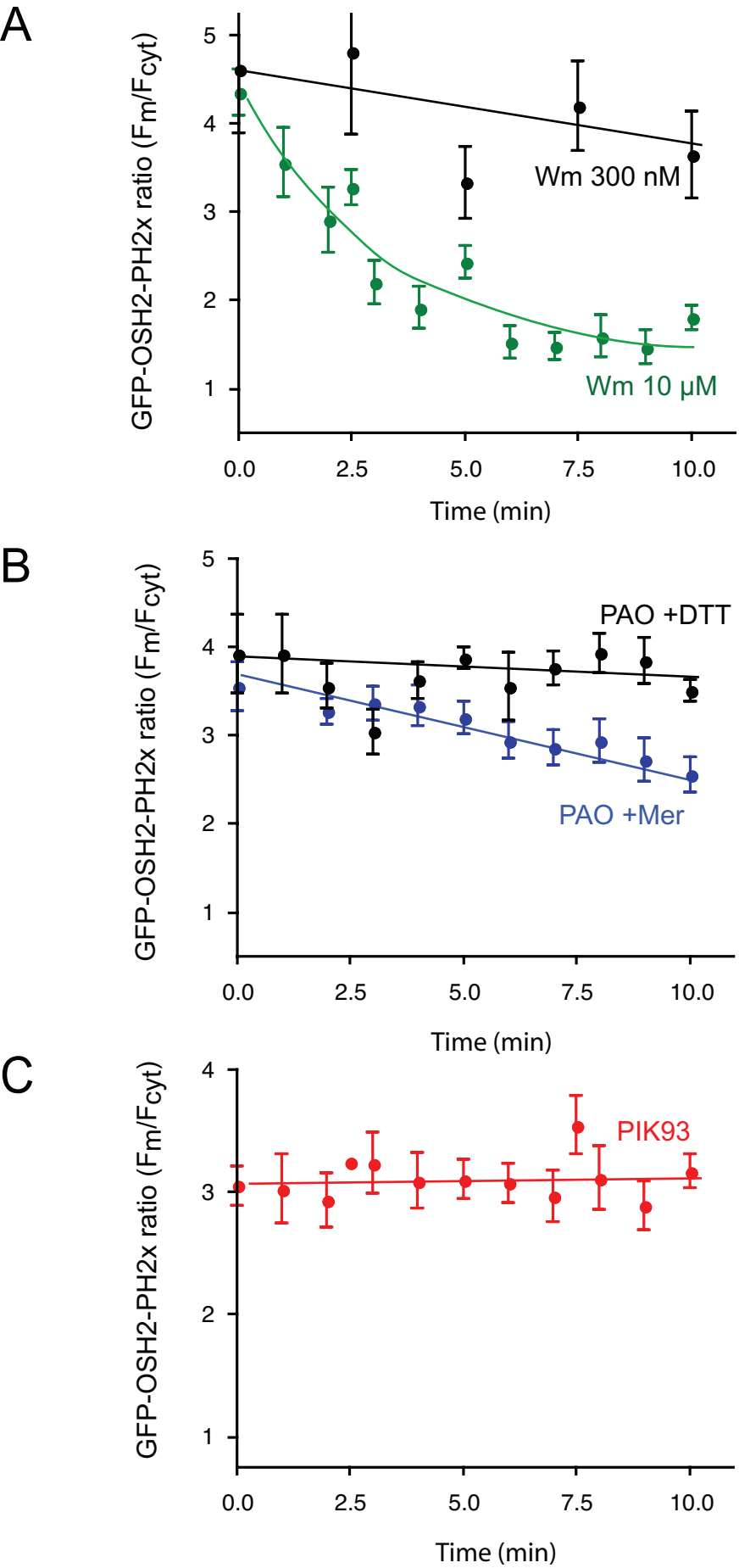
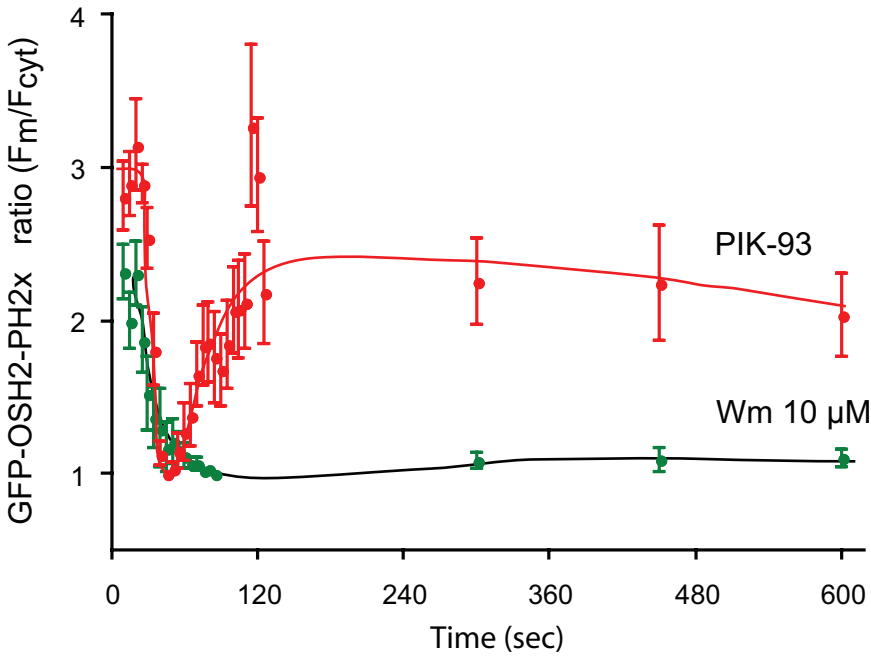

A
GFP-OSH2-PH2x ratio (Fm/Fcyt)
Wm 300 nM
Wm 10 µM
Time (min)
B
GFP-OSH2-PH2x ratio (Fm/Fcyt)
PAO +DTT
PAO +Mer
Time (min)
C
GFP-OSH2-PH2x ratio (Fm/Fcyt)
PIK93
Time (min)

Figure 7

A


GFP-OSH2-PH2x ratio (Fm/Fcyt)
4
3
2
1
0
120
240
360
480
600
Time (sec)
PIK-93
Wm 10 µM

B

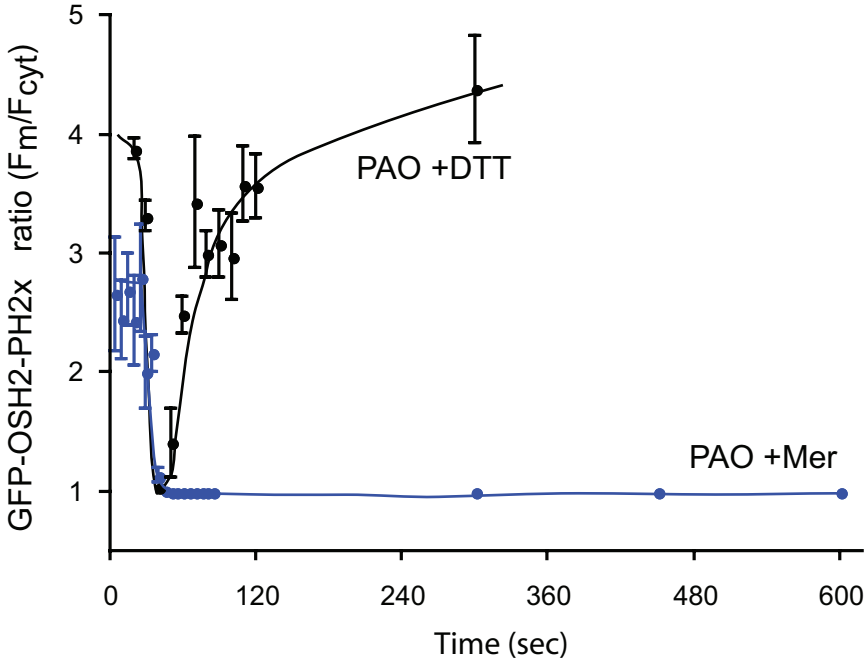

GFP-OSH2-PH2x ratio (Fm/Fcyt)
5
4
3
2
1
0
120
240
360
480
600
Time (sec)
PAO +DTT
PAO +Mer

Figure 8

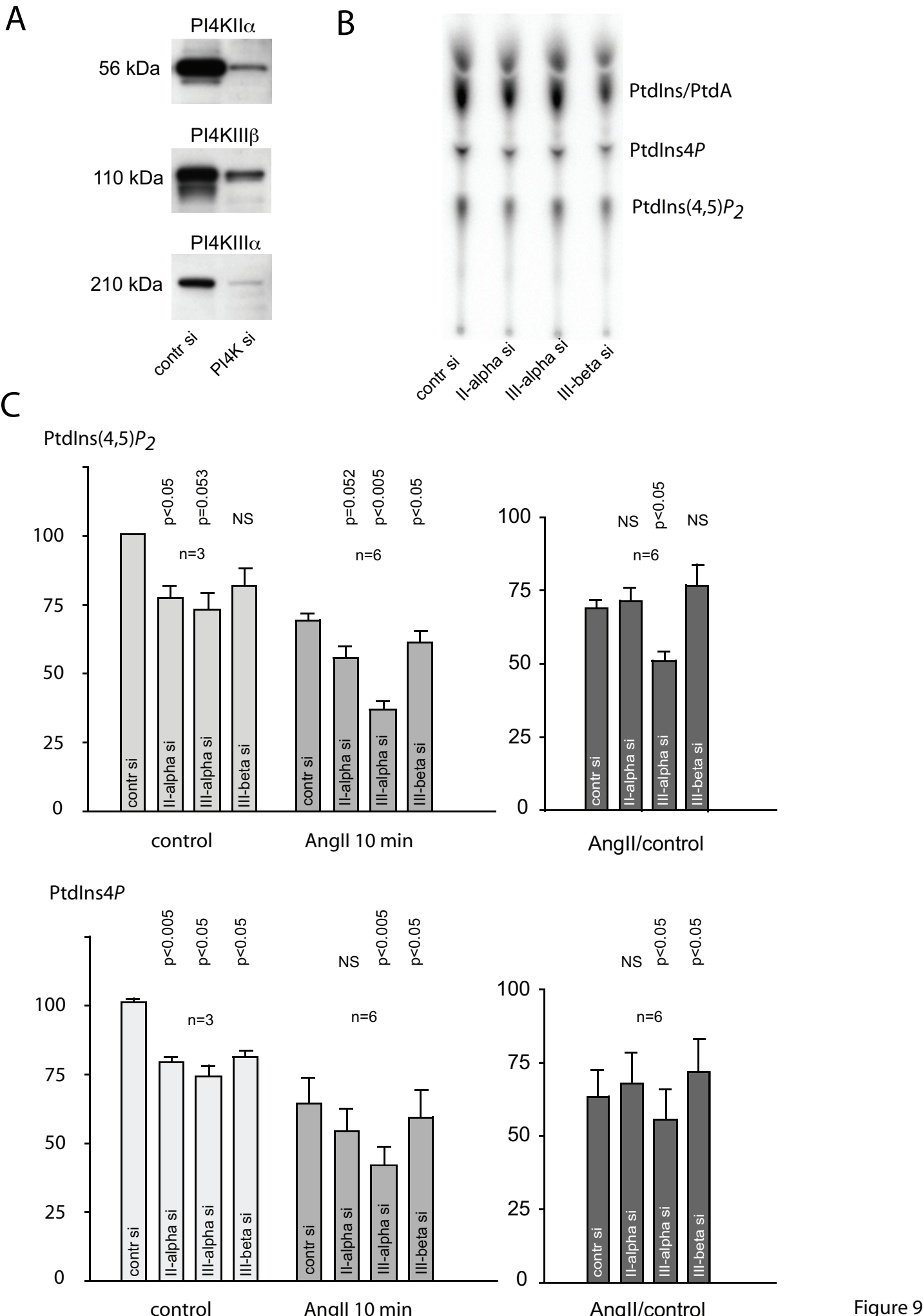

A
PI4KIIα
56 kDa
PI4KIIIβ
110 kDa
PI4KIIIα
210 kDa
contr si
PI4K si
B
PtdIns/PtdA
PtdIns4P
PtdIns(4,5)P2
contr si
II-alpha si
III-alpha si
III-beta si
C
PtdIns(4,5)P2
p<0.05
p=0.053
NS
p=0.052
p<0.005
p<0.05
n=3
n=6
100
75
50
25
0
contr si
II-alpha si
III-alpha si
III-beta si
control
AngII 10 min
NS
p<0.05
NS
n=6
AngII/control
PtdIns4P
p<0.005
p<0.05
p<0.05
NS
p<0.005
p<0.05
n=3
n=6
NS
p<0.05
p<0.05
n=6

Figure 9